Phylogenetic history, allometry and disparate functional pressures influence the morphological diversification of the gekkotan quadrate, a keystone cranial element

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The functional components of the vertebrate skull—including the chondrocranial braincase, protective dermatocranium and lower jaw—are interconnected and become operational through the quadrate bone. This latter element is critical for cranial biomechanics and support of the auditory system in squamate reptiles, but the interspecific variation of quadrate anatomy has not been studied in detail. Our objectives were to determine the relative influence of phylogenetic history, allometry and functional selection pressures on the morphological diversification of this keystone cranial element across gecko genera using high-resolution X-ray micro-computed tomography, three-dimensional geometric morphometrics and phylogenetic comparative methods. Our results demonstrate substantial variation in gecko quadrate morphology. Two families possess highly derived quadrate morphologies, while the remaining gekkotans retain extensive overlap in quadrate shape. Allometric scaling has influenced shape across species; distantly related miniaturized taxa possess elongate, slender quadrate morphologies while large taxa have robust, laterally expanded quadrates. The relative height of the coronoid eminence and the loss of the peripheral auditory system co-vary with quadrate anatomy, indicating that multiple disparate functional pressures may act on this element. Our study has identified the disparity of quadrate morphology within geckos and has highlighted the importance of considering multiple factors that may influence the diversification of phenotypes.

ADDITIONAL KEYWORDS: anatomy - computed tomography - cranial evolution - gecko - geometric morphometrics - skull.

INTRODUCTION

A long-standing challenge in evolutionary biology is the classification and interpretation of phenotypic diversity. Historically, researchers have experienced limitations in sampling taxa broadly, quantifying complex morphologies and disentangling the complex mechanisms responsible for generating macroevolutionary patterns. The diversification of phenotypes within a clade may be an adaptive response to ecological and functional pressures that can lead to divergence or convergence, depending on the selective regime (Losos, 1990; Grant & Grant, 2002). Phylogenetic and architectural constraints can have a restrictive influence, however, yielding phenotypic conservatism or allometric scaling of structures (Konstruktions-Morphologie; Seilacher, 1970; Gould & Lewontin,

1979). Furthermore, complex phenotypes, including the vertebrate skull, may be modular and partitioned into semi-indepedent blocks of traits, and these modules may be differentially influenced by historical, architectural and adaptive mechanisms (Drake & Klingenberg, 2010; Goswami & Polly, 2010; Felice & Goswami, 2017). By employing recent advances of micro-computed tomography, three-dimensional (3D) geometric morphometrics and phylogenetic comparative methods, we investigated the relative roles of phylogenetic history, allometry and functional pressures in the morphological diversification of a single unit of the lepidosaurian skull across the majority of gecko genera.

Gekkotans are one of the most successful vertebrate lineages, comprising more than 25% of all described lizard species. They are phenotypically diverse, varying greatly in body size, habitat and activity preferences, and diet (Daza *et al.*, 2009; Gamble *et al.*, 2012, 2015). Furthermore, the interrelationships of geckos are well understood and are highly supported in recent

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molecular phylogenies (Gamble *et al.*, 2012, 2015). The skull anatomy of geckos has been studied in multiple taxa (see Daza *et al.*, 2008: appendix), and a general reduction in the cranial bones occurs in nearly all species relative to the plesiomorphic squamate conditition (e.g. loss of the postorbital and supratemporal bars), although the paired quadrate bones remain prominent and central to skull structure (Fig. 1).

The functional units of the lizard skull are divided into the chondrocranial braincase, protective dermatocranium and lower jaw (Rieppel, 1993; Evans, 2008). These regions are interconnected and become operational through a fourth functional unit, the quadrate bone. The quadrate is a paired bone that is located posterolaterally in the skull, flanking either side of the braincase. The overall structure of the quadrate is complex and has been variously described as auricle- (Oelrich, 1956), conch- (Earle, 1961), shell-(Daza et al., 2008), lamina- (Evans, 2008) or even tacoshaped (J. D. Daza, pers. comm.). Generally, there is a medial column that supports dorsal (cephalic) and ventral (mandibular) condyles (Fig. 2). The lateral wing of the quadrate forms a posterior concavity (conch), and the midline of this structure supports the anterior edge of the tympanic membrane of the ear (Fig. 1), whereas the rest of the membrane circumference is supported by connective tissue (Wever, 1973). The stapes and extracolumella also possess either a direct contact or a soft-tissue connection with the quadrate. The cephalic condyle articulates with the parocciptial process of the otooccipital (chondrocranial braincase), as well as the supratemporal and squamosal (protective dermatocranium; although these two articulations are often lost in geckos), while the mandibular condyle articulates with the articular of the lower jaw. The medial surface has a reduced facet for articulation with

the pterygoid, dorsal to the mandibular condyle (Fig. 1). The anterior surface of the quadrate also acts as an attachment area for multiple cranial muscles, including the massive adductors, which function to close the jaw (Rieppel, 1984; Daza et al., 2011). Because of these many connections, the quadrate is an important functional unit for support of the peripheral auditory system and for cranial biomechanics. The quadrate also plays a critical role in squamate cranial kinesis: anteroposterior translation of this element, or streptostyly, pulls the palatal bones of the skull posteriorly, which depresses the muzzle bones at the mesokinetic joint (ventroflexion between the parietal and frontal; Versluys, 1912; Frazzetta, 1962). Multiple hypotheses exist regarding the function of squamate cranial kinesis (Metzger. 2002), and of these, improved prey manipulation is the most well supported (Rieppel, 1978).

The functional demands placed on the quadrate are likely particularly high in gekkotans, as these animals are known to exhibit pronounced hearing sensitivity (Werner et al., 2002, 2005) and cranial kinesis (Herrel et al., 1999, 2000, 2007). While this element has been morphologically described for a few individual gekkotan taxa, a large-scale, comparative analysis has yet to be undertaken. The objectives of this study were to answer three main questions: (1) Are there differences in the shape of the quadrate among the different family-level clades of geckos, indicating an influence of phylogenetic history? (2) Do similarly sized species show comparable patterns in quadrate morphology across clades, demonstrating architectural constraints related to allometry? (3) Do hypothesized functional traits vary among species relative to quadrate morphology, suggesting that quadrate shape changes in response to functional selection pressures? Answering these questions will allow us to identify the disparity of

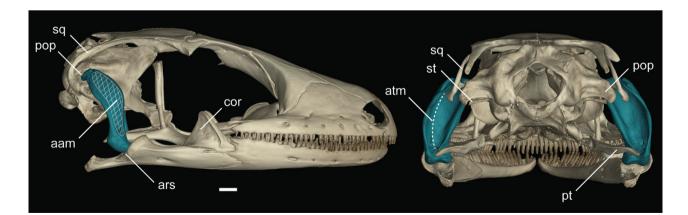


Figure 1. Skull of *Toropuku stephensi* (Diplodactylidae; CAS 47986) in lateral (left) and posterior (right) view. Quadrate is highlighted in blue. Abbreviations: aam, attachment area for adductor mandibulae muscles; ars, articular surface; atm, attachment area for tympanic membrane; cor, coronoid; pop, paroccipital process; pt, pterygoid; sq, squamosal; st, stapes. Scale bar = 1 mm.

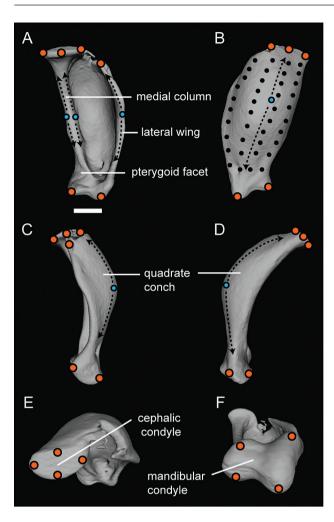


Figure 2. Quadrate of Toropuku stephensi (Diplodactylidae; CAS 47986) in posterior (A), anterior (B), lateral (C), medial (D), dorsal (E) and ventral (F) views. Three-dimensional landmarks digitized on quadrate shape files: nine fixed landmarks (orange points) that correspond to the cephalic condyle, mandibular condyle and dorsolateral edge of the lateral wing; four sliding semi-landmarks (blue points) that correspond to the maximal lateral edge of the quadrate conch, relative width of the medial column and anterior extent of the conch (dashed lines indicate the direction of the sliding semi-landmarks); and 40 surface semi-landmarks (black points) that correspond to the anterior surface of the quadrate conch. Scale bar = 1 mm.

quadrate morphology across geckos and better elucidate which processes have influenced the phenotypic diversification of this keystone cranial element.

MATERIAL AND METHODS

We investigated quadrate morphology within the Gekkota using high-resolution X-ray micro-computed tomography (CT). Specimens were scanned on an

Xradia MicroCT scanner (Pleasanton, CA, USA) at the University of Texas at Austin High Resolution X-ray CT-Facility (http://www.ctlab.geo.utexas.edu/). In total, 132 specimens representing all families and nearly all genera (116 of 125) were CT-scanned and included in our analyses (Supporting Information, Table S1). All individuals examined were believed to be adults although a few subadult specimens may be included due to intraspecific variation in size at sexual maturity - as we have demonstrated elsewhere that ontogeny, but not sexual dimorphism, drives the intraspecific variation of quadrate morphology in a single species of gecko (Paluh et al., 2018). Three-dimensional stereolithography (STL) files and CT settings are available to view and download from Duke University's morphological data archive (http://www.morphosource.org).

A 3D model of the right quadrate was generated for each specimen using the segmentation tools in the software Avizo v.9.1 (VSG, Visualization Sciences Group, Burlington, MA, USA) and exported in polygon file format (PLY) for subsequent analyses. Three-dimensional geometric morphometric analyses were used to quantify variation in quadrate shape using the R package geomorph v.3.0.3 (Adams & Otarola-Castillo, 2013). Nine fixed landmarks, four sliding semi-landmarks and 40 surface semi-landmarks were digitized on each quadrate using geomorph. The fixed landmarks correspond to homologous and repeatable points of the quadrate: the cephalic condyle, mandibular condyle and dorsolateral edge of the lateral wing; the sliding semi-landmarks correspond to the medial column, maximal lateral edge of the lateral wing and maximal anterior extent of the conch; and the surface semi-landmarks correspond to the anterior surface of the quadrate conch (Fig. 2). A generalized Procrustes analysis was performed to align, rotate and scale specimens to a common coordinate system and unit-centroid size to remove variation in their position, orientation and size (Rohlf & Slice, 1990), resulting in a set of shape variables, or Procustes tangent coordinates, for all specimens.

PHYLOGENETIC HISTORY

To visualize quadrate shape variation among all gecko species examined, we performed a principal component analysis (PCA) using the Procustes tangent coordinates, and the Procrustes-aligned specimens were plotted in two dimensions of tangent space (PC1 and PC2) and grouped by clade membership. A multilocus, genus-level molecular phylogeny for Gekkota (Gamble *et al.*, 2015) was pruned to correspond to the taxa in the shape variables data set and provided an estimate of the evolutionary relationships. Head length measurements and principal component scores were mapped onto the phylogeny using the R package phytools (Revell, 2012). We calculated the *K*-statistic's

generalization for multivariate data (Kmult; Adams, 2014a) to determine the strength of phylogenetic signal in quadrate shape. The P-value evaluates the magnitude of phylogenetic signal in the set of shape variables and the Kmult statistic assesses the fit of a Brownian motion (BM) evolutionary model of trait evolution (Blomberg et al., 2003). The magnitude of phylogenetic signal was evaluated for all taxa combined (Gekkota), as well as for each familial clade separately. To visualize the degree of phylogenetic signal, the gekkotan phylogeny was projected into quadrate morphospace by calculating ancestral states of the internal nodes through maximum likelihood, as implemented in geomorph. Analysis of covariance (ANCOVA; Collyer et al., 2015) and phylogenetic least squares (PGLS; Adams, 2014b) tests were performed using the aligned Procustes tangent coordinates to determine whether the seven families of geckos occupy different regions of quadrate morphospace (Table 1). Post-hoc pairwise comparisons of group means were calculated to determine which gekkotan clades have diverged from one another in quadrate morphology.

ALLOMETRY

We examined the influence of allometry by assessing the multivariate and phylogenetic regression between quadrate shape and quadrate centroid size (CS; Bookstein, 1991; method RegScore) based on Procrustes distances. A homogeneity of slopes (HOS) test and post-hoc pairwise slope comparisons were performed to determine if the seven families of geckos have diverged from one another in the allometric relationship of quadrate shape to size. A regression plot was generated to visualize the multivariate relationship between size and shape of gekkotan quadrates using 'RegScore' shape scores (Drake & Klingenberg, 2008) and ordinary least squares regression lines were generated for each family to visualize differences in slope. We further compared the relationship between quadrate size (centroid size) and quadrate shape (Regscore) to linear measurements of the head (log-transformed length, width and depth). Lastly, the interaction of quadrate centroid size and the main effects of the other ANCOVAs and PGLSs (clade membership, coronoid height, head depth, external ear presence) were also tested (Table 1).

FUNCTIONAL ASSOCIATIONS

Due to the lack of ecological (e.g. diet) and biomechanical (e.g. bite force) data available for geckos across all genera, we tested if quantifiable hypothesized functional traits co-vary among species relative to quadrate shape. Previous research has demonstrated that relative head depth and relative coronoid eminence

height may be a strong predictor of bite force and diet across many squamate groups (Herrel et al., 2001; Herrel et al., 2004; McBrayer, 2004; Metzger & Herrel, 2005; Lappin et al., 2006), as skull height influences the orientation and length of the jaw adductors (which influences torque) and coronoid height corresponds to the available insertion area for the jaw adductors (Daza et al., 2011). We measured head depth, coronoid height and head length of all taxa using the CT data and measurement tools in VGStudio Max 3.0 (Volume Graphics, Heidelberg, Germany). Both head depth and coronoid height were first divided by head length to remove the influence of overall skull size. An ANCOVA and PGLS were performed to determine whether a predicted relationship exists between quadrate shape and relative head depth and between quadrate shape and relative coronoid height (Table 1).

To determine whether a predicted relationship exists between quadrate shape and the peripheral auditory system of geckos, we conducted a survey of the presence/absence of the external ear system (external auditory meatus and tympanum) through examination of specimen photographs and transverse cross-sections of the X-ray tomograms. The pygopodids in the genus Aprasia have been previously described as possessing a rudimentary auditory system, lacking an external ear, tympanum and stapes (Shute & Bellairs, 1953; Manley & Kraus, 2010; Daza & Bauer, 2015), but this has yet to be reported in other gekkotans. An ANCOVA and PGLS were performed between gekkotans that possess the external auditory system and those that lack these structures (Table 1).

RESULTS

Substantial shape variation was observed across the 132 gekkotan quadrates we examined (Fig. 3). The carphodactylids possess a relatively large and robust quadrate that has a laterally expanded wing, wide medial column and expanded condyles (Fig. 3A–D), while the pygopodids have dorsoventrally inflected quadrates that contain either a large posterior concavity (Fig. 3F, H) or a bilaterally compressed column-like structure (Fig. 3E, G, I). The remaining five families possess quadrate shapes that are less distinct than the Carphodactylidae and Pygopodidae but vary in relative size of the cephalic and mandibular condyles, width of the medial column and lateral wing, and size of the posterior concavity (Fig. 3J-AR). The diversity of quadrate shapes observed in the Eublepharidae (Fig. 3S-W) and Phyllodactylidae (Fig. 3AD-AH) is lower than that in the Diplodactylidae (Fig. 3J-R), Sphaerodactylidae (Fig. 3X-AC) and Gekkonidae (Fig. 3AI-AR). The first two axes of the PCA explain 43% of the total shape variation observed in the

Table 1. Results from ANCOVA and PGLS tests comparing trends in quadrate shape divergence using familial clades, coronoid height, head depth and ear presence as the main effects, centroid size as the co-variate, and the interaction between each main effect and centroid size

Model	d.f.	SS	MS	R^2	F	Z	P value
Multivariation regression							
Centroid size	1	0.2114	0.21140	0.16103	24.952	7.3725	< 0.001
Phylogenetic regression							
Centroid size	1	0.8210	0.82105	0.088493	12.621	5.3333	< 0.001
ANCOVA (clade)							
Clade	6	0.28211	0.047019	0.214892	6.8294	4.5685	< 0.001
Centroid size	1	0.12480	0.124802	0.095065	18.1272	14.0408	< 0.001
Clade:centroid size	6	0.09349	0.015582	0.071213	2.2632	2.2093	< 0.001
Residuals	118	0.8124	0.006885				
Total	131	1.3128					
PGLS (clade)							
Clade	6	0.3015	0.05025	0.032496	0.7991	7.1322	< 0.001
Centroid size	1	0.7451	0.74507	0.080304	11.8492	12.3826	< 0.001
Clade:centroid size	6	0.8118	0.1353	0.087497	2.1518	3.2744	< 0.001
Residuals	118	7.4198	0.06288				
Total	131	9.2781					
ANCOVA (coronoid)							
Coronoid	1	0.09350	0.093497	0.071219	11.0621	8.2875	< 0.001
Centroid size	1	0.114835	0.114835	0.087473	13.5866	10.9951	< 0.001
Coronoid:centroid size	1	0.02261	0.02261	0.017223	2.6751	2.4515	0.008
Residuals	128	1.08186	0.008452				
Total	131	1.3128					
PGLS (coronoid)							
Coronoid	1	0.3586	0.35855	0.038645	5.6412	2.702	0.003
Centroid size	1	0.5768	0.57678	0.062116	9.0747	8.6154	< 0.001
Coronoid:centroid size	1	0.2072	0.20716	0.022327	3.2593	1.3188	0.119
Residuals	128	8.1357	0.06356				
Total	131	9.2781					
ANCOVA (head depth)							
Head depth	1	0.01670	0.016698	0.012719	1.9545	1.4823	0.098
Centroid size	1	0.19222	0.192224	0.146422	22.5003	7.0006	< 0.001
Head depth:centroid size	1	0.01036	0.010357	0.00789	1.2124	1.118	0.24
Residuals	128	1.09353	0.008543				
Total	131	1.3128					
PGLS (head depth)							
Head depth	1	0.596	0.59596	0.064233	10.579	0.3719	0.955
Centroid size	1	0.8235	0.82351	0.088758	14.618	5.8346	< 0.001
Head depth:centroid size	1	0.648	0.64799	0.069841	11.503	0.495	0.926
Residuals	128	7.2107	0.05633				***-*
Total	131	9.2781	0.0000				
ANCOVA (ear)	101	0.2.01					
Ear	1	0.12362	0.123621	0.094165	15.9039	10.0102	< 0.001
Centroid size	1	0.18551	0.185508	0.141307	23.8658	17.6793	<0.001
Ear:centroid size	1	0.00873	0.008734	0.006653	1.1237	0.9857	0.275
Residuals	128	0.99494	0.007773	0.000000	1.1201	0.5001	0.210
Total	131	1.3128	0.001110				
PGLS (ear)	101	1.0120					
Ear	1	0.7015	0.70152	0.075610	11.6539	8.2447	<0.001
Centroid size	1	0.7513	0.75100	0.080835	12.4592	12.1547	<0.001
Ear:centroid size	1	0.1215	0.12151	0.013096	2.0186	1.5736	0.089
Residuals	128	7.7051	0.12151	0.019090	2.0100	1.9790	0.009
nesiduais	140	1.1001	0.0004				

Boldfaced values indicate statistical significance.

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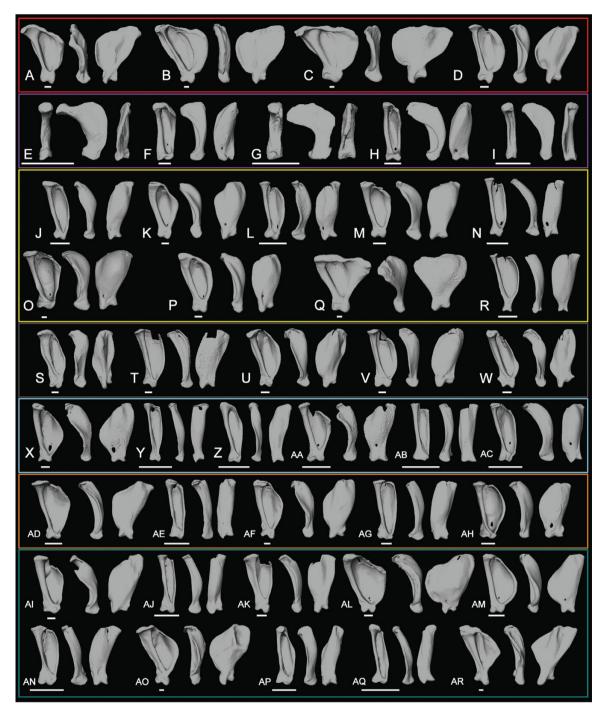


Figure 3. Representative quadrates from the seven families of geckos in posterior, lateral and anterior views from left to right. Carphodactylidae: A, Carphodactylus laevis (MCZ R-35114); B, Nephrurus asper (CAS 74733); C, Saltuarius cornutus (FMNH 57503); D, Underwoodisaurus milii (CAS 74744). Pygopodidae: E, Aprasia repens (CAS 104382); F, Lialis burtonis (FMNH 166958); G, Ophidiocephalus taeniatus (AMS R45179); H, Paradelma orientalis (CAS 77652); I, Pletholax gracilis (MCZ R-187676). Diplodactylidae: J, Amalosia rhombifer (CAS 100919); K, Correlophus belepensis (CAS 250865); L, Crenadactylus ocellatus (CAS 95287); M, Dactylocnemis pacificus (CAS 47979); N, Dierogekko insularis (AMS R161070); O, Hoplodactylus duvaucelii (CM 51270); P, Mniarogekko jalu (CAS 250858); Q, Rhacodactylus auriculatus (CAS 205486); R, Rhynchoedura ornata (UMMZ 124484). Eublepharidae: S, Aeluroscalabotes felinus (FMNH 146141); T, Eublepharis macularius (CM 67524); U, Goniurosaurus kuroiwae (CAS 198810); V, Hemitheconyx caudicinctus (CAS 165588); W, Holodactylus africanus (CAS 198932). Sphaerodactylidae: X, Aristelliger georgeensis (CAS 176485); Y, Coleodactylus brachystoma (UMMZ

quadrate across geckos (PC1 = 30% and PC2 = 13%). The PC1 axis describes shape differences driven by the relative expansion of the quadrate lateral wing: species with a negative PC1 score possess broad quadrates that have large lateral expansions while species with a positive PC1 score have narrow quadrates and no lateral expansion. The PC2 axis describes the shape differences driven by the degree of quadrate inflection: taxa with a negative PC2 score possess vertical quadrates and species with a positive PC2 score have dorsoventrally inflected quadrates.

PHYLOGENETIC HISTORY

Mapping these principal component scores onto the phylogeny of Gamble et al. (2015) indicates that there is generally little correspondence between the direction of PC values and clade membership, with the exception of the Carphodactylidae (red clade; Fig. 4A) and Pygopodidae (purple clade; Fig. 4A). The PCA plot further demonstrates that the gekkotan families are largely overlapping in morphospace, except for the Pygopodidae and Carphodactylidae, which are separated by the PC2 axis and PC1 axis, respectively (Fig. 4B). The pygopodids possess a dorsoventrally inflected quadrate, and the carphodactylids possess a robust, vertical quadrate that is characterized by a laterally expanded wing. The ANCOVA and PGLS indicated that quadrate shape divergence across the gekkotan families is significant (Table 1) and post-hoc pairwise comparisons of group means indicate that the Carphodactylidae and Pygopodidae possess derived quadrates, while the quadrate shapes of the remaining five families are not different from one another (Table 2). The K-statistic's generalization for multivariate data suggested that less phylogenetic signal is present than expected under a BM evolutionary model of trait evolution for Gekkota as a whole and the individual familial clades, although the Carphodactylidae and Eublepharidae approach significance (Table S2, Fig. S1).

ALLOMETRY

The multivariate regression and phylogenetic regression between quadrate shape and centroid size exhibited significant allometry, indicating there is a quadrate shape to size relationship within the Gekkota (Table 1; Fig. 4C). In general, small geckos

have narrow, slender quadrates and large geckos have robust, laterally expanded quadrates (Fig. 5). The HOS test suggested that the gekkotan families display differing patterns of allometry and post-hoc pairwise slope comparisons indicate that the Carphodactylidae have a distinct slope vector length from all other families, whereas the Pygopodidae have a different slope vector orientation compared to four of the families (Table 3; Fig. 4C). The linear measurements of the head (log-transformed length, width and depth) were highly correlated with one another across all specimens in this study $(R^2 > 0.9)$, and quadrate centroid size was highly correlated with these linear measurements ($R^2 > 0.9$; Fig. 6). This is in contrast to quadrate shape (Regscore), which was only weakly correlated with the head measurements ($R^2 < 0.7$; Fig. 6). A significant interaction between centroid size and clade membership was identified in both the ANCOVA and the PGLS (Table 1).

FUNCTIONAL ASSOCIATIONS

A predicted relationship was found between quadrate shape and size-corrected coronoid height (Table 1), suggesting that the morphology of the quadrate and coronoid may co-vary, independent of size, in response to functional selection pressures related to bite force and diet. As the relative height of the coronoid increases, the relative width of the quadrate increases. A significant interaction between centroid size and coronoid height was identified in the ANCOVA, but not in the PGLS. The size-corrected head depth ANCOVA and PGLS indicated that no relationship exists between quadrate shape variation and relative head depth variation (Table 1). External ears, tympana and stapes were present in all gekkotans, with the exception of three miniaturized pygopodid genera. Aprasia repens lacks all three of these structures, whereas Ophidiocephalus taeniatus and Pletholax gracilis possess a reduced stapes (footplate only) and lack a tympanum and external auditory meatus. The ANCOVA and PGLS indicated that quadrate shape divergence between nontympanic pygopodids and the remaining gekkotans is significant (Table 1). The non-tympanic pygopodids possess dorsoventrally inflected, columnar quadrates that are mediolaterally compressed and lack a posterior concavity and lateral wing (Fig. 3E, G, I).

103051); Z, Lepidoblepharis xanthostigma (CAS 178104); AA, Pristurus carteri (CAS 225349); AB, Saurodactylus fasciatus (CAS 92404); AC, Sphaerodactylus semasiops (MCZ R-55766). Phyllodactylidae: AD, Asaccus elisae (CAS 218137A); AE, Garthia gaudichaudii (UMMZ 111574); AF, Haemodracon riebeckii (MCZ A-27255); AG, Phyllodactylus baurii (CAS 9501); AH, Tarentola mauritanica (CAS 87112). Gekkonidae: AI, Ailuronyx seychellensis (CAS 167459); AJ, Alsophylax pipiens (CAS 143679); AK, Calodactylodes aureus (MCZ R-3918); AL, Chondrodactylus bibronii (CAS 173299); AM, Crossobamon eversmanni (CAS 180001); AN, Ebenavia inunguis (CAS 66195); AO, Gekko gecko (SHSVM-H-0001-2014); AP, Hemiphyllodactylus typus (CAS 174223); AQ, Microgecko helenae (CAS 120795); AR, Uroplatus fimbriatus (CAS-SU 13469). Scale bars = 1 mm.

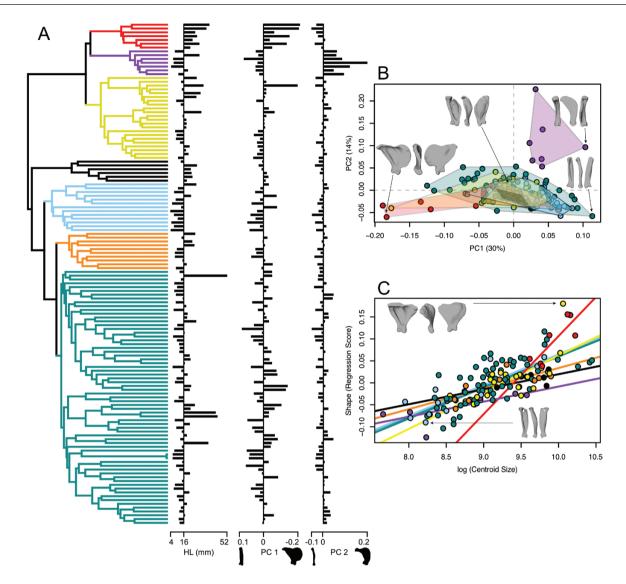


Figure 4. A, Head length (HL, mm) and quadrate principal component scores (PC1 and PC2) of all specimens mapped onto the genus-level phylogeny of Gamble et al. (2015) demonstrating little corresponse between clade membership and the direction of PC values, with the exception of the Carphodactylidae and Pygopodidae. Family-level clades are colour-coded: red = Carphodactylidae, purple = Pygopodidae, yellow = Diplodactylidae, black = Eublepharidae, blue = Sphaerodactylidae, orange = Phyllodactylidae, cyan = Gekkonidae. B, PCA plot of shape variation exhibiting the diversity of quadrate morphologies within geckos. Divergent quadrates are shown in posterior (left), lateral (centre) and anterior (right) views: Saltuarius salebrosus (CAS 74742; Carphodactylidae; bottom left), Naultinus elegans (CAS 47976; Diplodactylidae; centre), Pletholax orientalis (MCZ R-187676; Pygopodidae; top right), Matoatoa breviceps (MCZ R-190009; Gekkonidae; bottom right). C, multivariate regression between quadrate shape (RegScore) and quadrate centroid size. Ordinary least squares regression lines are displayed for each family to demonstrate the slope differences in Carphodactylidae and Pygopodidae. Quadrates are shown from a large species (Rhacodactylus auriculatus; CAS 205486; Diplodactylidae; top right) and a small species (Coleodactylus brachystoma; UMMZ 103051; Sphaerodactylidae; bottom left).

DISCUSSION

Through the use of micro-CT, 3D geometric morphometrics and phylogenetic comparative methods, we were able to begin disentangling the complex mechanisms responsible for generating the phenotypic diversity in a single

functional unit of the gecko skull. Our results indicate that phylogenetic history, allometry and functional demands have differentially influenced the diversification of gekkotan quadrate morphology. We discuss the results for our three main questions in further detail below.

 $\textbf{Table 2.} \ \textit{P-} \textbf{values for pairwise distances of group means associated with ANCOVA examining quadrate shape differences across gecko families$

	Carph	Diplo	Euble	Gekk	Phyllo	Pygo	Sphaer
Carph	1	*	*	*	*	*	*
Diplo	< 0.01	1	*	*	*	*	*
Euble	< 0.01	0.1	1	*	*	*	*
Gekk	< 0.01	0.09	0.3	1	*	*	*
Phyllo	< 0.01	0.5	0.3	0.8	1	*	*
Pygo	< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	1	*
Sphaer	<0.01	0.06	0.2	0.2	0.6	<0.01	1

Carph, Carphodactylidae; Diplo, Diplodactylidae; Euble, Eublepharidae; Gekk, Gekkonidae; Phyllo, Phyllodactylidae; Pygo, Pygopodidae; Sphaer, Sphaerodactylidae.

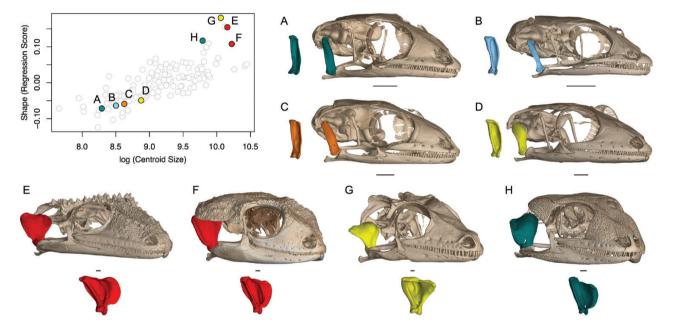


Figure 5. A–D, four miniaturized taxa from different families that possess elongate, slender quadrates: A, $Microgecko\ helenae\ [CAS\ 120795; Gekkonidae; 6.05\ mm\ head\ length\ (HL); B, <math>Pseudogonatodes\ barbouri\ (MCZR\ 14385; Sphaerodactylidae; 4.24\ mm\ HL); C, <math>Garthia\ gaudichaudii\ (UMMZ\ 111574; Phyllodactylidae; 7.88\ mm\ HL); D, <math>Amalosia\ rhombifer\ (CAS\ 100919; Diplodactylidae; 10.61\ mm\ HL).\ E–H, four\ large\ taxa\ that\ possess\ robust,\ laterally\ expanded\ quadrates: E, <math>Saltuarius\ cornutus\ (FMNH\ 57503; Carphodactylidae; 37.38\ mm\ HL); F, <math>Nephrurus\ asper\ (CAS\ 74733; Carphodactylidae; 28.77\ mm\ HL); G, Rhacodactylus\ auriculatus\ (CAS\ 205486; Diplodactylidae; 32.27\ mm\ HL); H, <math>Chondrodactylus\ bibronii\ (CAS\ 173299; Gekkonidae; 21.63\ mm\ HL).$ Inset: multivariate regression between quadrate shape and quadrate centroid size highlighting taxa\ A–H. See colour\ designations\ of\ Figure\ 4. Scale bars = 1 mm.

ARE THERE QUADRATE SHAPE DIFFERENCES AMONG DIFFERENT CLADES OF GECKOS?

The Pygopodidae and Carphodactylidae occupy unique areas of quadrate morphospace and are significantly divergent from other gekkotans in shape, suggesting that the quadrate structure of these two families are derived and have been influenced by their respective phylogenetic histories. All pygopodids possess a dorsoventrally inflected quadrate (Fig. 3E–I), while all carphodactylids possess a robust, vertical quadrate that is characterized by a laterally expanded crest

(Fig. 3A–D). Interestingly, these two families are sister lineages that are endemic to Australia (with one species of pygopodid in New Guinea), and both clades have also been shown to have an enlarged squamosal that participates in the formation of the postemporal bar and may contact the quadrate more strongly (Stephenson, 1960; Bauer, 1986; Daza & Bauer, 2012). Phylogenetic signal was absent or less than expected under BM for Gekkota as a whole, as well as within each family. As might be expected, particularly given our dense sampling at the generic level, clades with

Table 3. *P*-values for pairwise slope comparisons associated with homogeneity of slopes tests examining quadrate allometry differences across gecko families; the first set of *P*-values is for differences in slope vector length (magnitude) and the second set is for slope vector orientation differences

	Carph	Diplo	Euble	Gekk	Phyllo	Pygo	Sphaer
Carph	1	*	*	*	*	*	*
Diplo	< 0.01	1	*	*	*	*	*
Euble	< 0.01	0.98	1	*	*	*	*
Gekk	< 0.01	0.20	0.99	1	*	*	*
Phyllo	< 0.01	0.22	0.87	0.75	1	*	*
Pygo	0.01	0.21	0.84	0.02	0.02	1	*
Sphaer	< 0.01	0.70	0.99	0.36	0.36	0.11	1
Carph	1	*	*	*	*	*	*
Diplo	0.31	1	*	*	*	*	*
Euble	0.17	0.63	1	*	*	*	*
Gekk	0.57	0.22	032	1	*	*	*
Phyllo	0.23	0.50	0.52	0.62	1	*	*
Pygo	0.14	0.02	0.24	<0.01	<0.01	1	*
Sphaer	0.54	0.24	0.33	0.36	0.32	<0.01	1

Carph, Carphodactylidae; Diplo, Diplodactylidae; Euble, Eublepharidae; Gekk, Gekkonidae; Phyllo, Phyllodactylidae; Pygo, Pygopodidae; Sphaer, Sphaerodactylidae.

the most genera and sampled specimens (Gekkonidae, Diplodactylidae) occupy a larger amount of tangent space (convex hull area > 0.01) than the less speciose clades (convex hull area < 0.008) within the PCA plot (Fig. 4B).

Daza et al. (2009) conducted a 2D geometric morphometric analysis of the entire gekkotan skull, and similarly determined that pygopodids occupy a divergent region of morphospace due to lateral and dorsoventral compression of the skull. The disparity between pygopodids and other geckos in cranial anatomy may be due to many factors; however, the most likely influence is the possession of a highly derived, limb-reduced, serpentine form. Despite this extreme morphological modification, the Pygopodidae vary considerably in habitat specificity (Underwood, 1957; Gans, 1986), jaw mechanics (Patchell & Shine, 1986a), diet and foraging behaviour (Shine, 1986; Patchell & Shine, 1986b), and body size (Meiri, 2008). Therefore, we hypothesize that limb loss in this group was coupled with modifications to the skull as a whole, as well as to the quadrate specifically. Although subsequent morphological, functional and ecological diversification events took place, the Pygopodidae retain a quadrate morphology that is more similar among these species than to other gekkotans. The quadrates of the Carphodactylidae occupy a divergent region of morphospace that is characterized by a robust cephalic condyle, mandibular condyle and medial column, as well as a laterally expanded wing. We hypothesize that the disparity between carphodactylids and other gekkotans in quadrate morphology has been influenced by their phylogenetic history in relation to

both unique allometric scaling and cranial function related to foraging ecology. Members of this family are characterized by possessing relatively large bodies and disproportionately large, co-ossified heads (Bauer, 1986). It is likely that the robustness of the condyles and medial column are coupled with increases in ossification of the entire skull. Carphodactylids lack toepads, yet they have diversified into terrestrial, arboreal and rock outcrop habitat specialists (Cogger, 2014). Despite this diversity in habitat use, members of the Carphodacylidae retain similar dietary preferences in that they incorporate robust prey into their diet, including large scorpions, centipedes, cockroaches and even other lizards (Bauer, 1986, 1990a; How et al., 1990; Doughty & Shine, 1995). The flaring lateral crest in this group is likely utilized to increase the insertion area available for cranial musculature, which would permit an increased bite force and ability to consume large prey. This is in contrast to most geckos, which typically forage on small-bodied insect groups and spiders and rarely take larger prey (Daza et al., 2009). An alternative hypothesis is that the expanded lateral wing may be used to enhance sound detection, as for the pinna in mammals (Webster, 1966); however, auditory detection capabilities in carphodactylid geckos are completely unknown (Rohtla, 2016) but warrant investigation.

DO SIMILARLY SIZED SPECIES SHOW SIMILAR MORPHOLOGICAL PATTERNS ACROSS CLADES?

Our results indicate that gekkotan quadrate morphology is influenced by allometric scaling, as a positive relationship was identified between quadrate

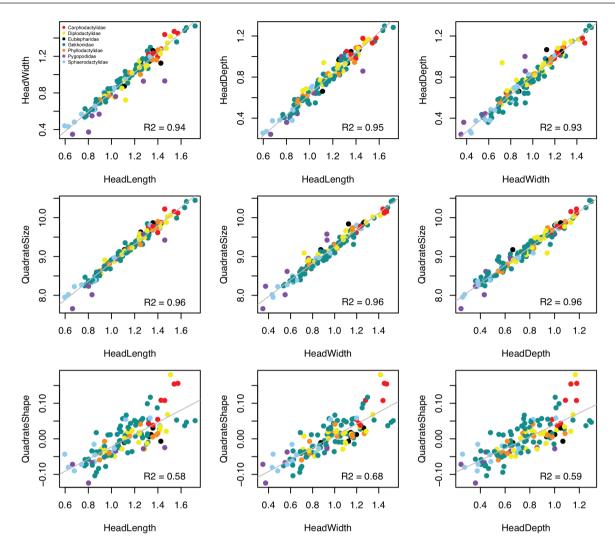


Figure 6. Regression relationships and ordinary least squares regression lines between: log-transformed head length, width and depth (top row), quadrate size (log-transformed centroid size) and log-transformed linear measurements of the head (middle row), and quadrate shape (RegScore) and log-transformed linear measurements of the head (bottom row).

shape and centroid size. The Carphodactylidae and Pygopodidae display differing patterns of allometry (Table 3, Fig. 4C), which may further explain the unique quadrate shape of these two families. We have previously shown that intraspecific quadrate shape variation in the gekkonid Hemidactylus turcicus is driven largely by allometric and ontogenetic change (Paluh et al., 2018). The direction of shape change through ontogeny in *H. turcicus* is similar to the general pattern we have identified across differently sized species: small geckos have narrow, slender quadrates while large geckos possess laterally expanded quadrates (Fig. 5). The amount of quadrate shape diversity across all species examined in this study (in specimens ranging from 4.0 to 52.6 mm head length) is much greater than the shape change observed during the development of *H. turcicus* (in specimens ranging

from 8.9 to 16.4 mm head length; Paluh et al., 2018). Most species in our current analyses are represented by one individual due to the broad taxonomic sampling across gekkotan genera; therefore, further work is needed to verify that ontogeny is the primary influence on intraspecific variation in quadrate morphology across other gecko species. The influence of allometric scaling on overall head shape has been previously identified in other lizards, including Anolis (Sanger et al., 2011) and Varanus (Openshaw & Keogh, 2014). Furthermore, allometric and ontogenetic scaling have been identified in lizard functional traits supported by the quadrate, including auditory sensitivity (Werner & Igíc, 2002; Werner et al., 2002, 2005) and bite force (Meyers et al., 2002; Verwaijen et al., 2002; Herrel & O'Reilly, 2006). Multiple studies have also identified ontogenetic variation in the diets of lizards, in which

juveniles eat smaller and softer prey than adults (Saenz, 1996; Angelici et al., 1997; Herrel et al., 2006; Whitfield & Donnelly, 2006). If allometric scaling in bite force exists across geckos due to diet variation, this may explain the significant association found between quadrate shape and the interaction of relative coronoid height and centroid size (Table 1). The relationship between quadrate centroid size and head dimensions is stronger than the association between quadrate shape and head dimensions (Fig. 6), indicating that head size alone can largely predict quadrate size, but that other factors are influencing quadrate shape. For example, there are miniaturized gecko genera across multiple families that possess a slender quadrate structure that lacks a laterally expanded wing, but variation remains between these taxa, including the relative size of the conch, cephalic condyle and mandibular condyle (Fig. 5A-D).

DO FUNCTIONAL TRAITS VARY AMONG SPECIES RELATIVE TO QUADRATE SHAPE?

A significant association was found between quadrate shape and relative coronoid height, suggesting that variation in quadrate morphology may be a response to selection pressures related to bite force and diet. The most extreme carphodactylid quadrates occupy a region of morphospace that is shared with one diplodactylid and two gekkonids that have converged on similar morphologies (Fig. 4B), including a large coronoid eminence (Fig. 5E-H). These taxa are also characterized by having disproportionately large heads and large body sizes. Rhacodactylus auriculatus (Diplodactylidae) has an extremely large quadrate lateral wing that flares anterolaterally into a triangular shape (Bauer, 1990b) and is very distant from any other diplodactylids in morphospace (Figs 3Q, 4B). Similar to the carphodactylids, multiple reports indicate that large prey items, including other lizards, are a substantial part of the R. auriculatus diet (Bauer & Sadlier, 1994; Snyder et al., 2010). Chondrodactylus angulifer and C. bibronii (Gekkonidae) possess a lateral expansion that is rounded (Figs 3AL, 5H), and previous work has documented that these taxa also commonly forage on geckos and other large prey (Loveridge, 1947; Pianka & Huey, 1978). These data suggest that the unique quadrate morphologies of the Carphodactylidae, Rhacodactylus and Chondrodactylus may be the result of unique cranial functional demands. Although significant, the relationship between quadrate shape and relative coronoid height was weak (Table 1), and no relationship was found between quadrate shape and head depth. To better investigate if variation in quadrate morphology is driven by cranial biomechanial pressures, we suggest future work

quantifies cranial myology and bite force variation, as well as diet variation, across gekkotan genera.

A unique pattern was identified within the Pygopodidae in relation to quadrate morphology and the presence/absence of the external auditory system elements. Four surface-dwelling species (*Delma borea*, *Lialis burtonis*, *Paradelma orientalis* and *Pygopus lepidopodus*) possessed an external ear, tympanic membrane and fully formed stapes, while the remaining three fossorial species (*Aprasia repens*, *Ophidiocephalus taeniatus* and *Pletholax gracilis*) lacked the external ear and tympanum. The stapes and extracolumella are absent in *Aprasia* (Daza & Bauer, 2015) and are highly reduced in *Pletholax* and *Ophidiocephalus* (Figs 7A, S2). The rudimentary ear and derived quadrate morphology of *Aprasia* have previously been described (Shute &

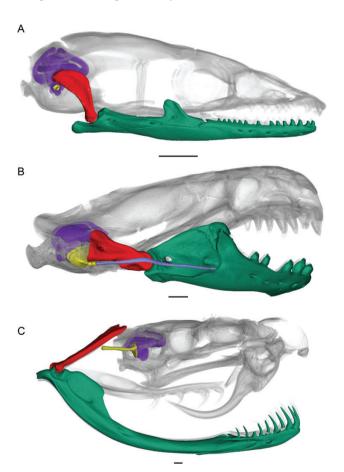


Figure 7. Three squamate groups that may use non-tympanic reception of substrate vibrations using divergent structural pathways between the lower jaw and inner ear: A, *Pletholax orientalis* (MCZ R-187676; Pygopodidae); B, *Amphisbaena fuliginosa* (UF 63167; Amphisbaenia); C, *Crotalus adamanteus* (UF 103268; Serpentes). Purple = inner ear endocast, yellow = stapes, blue = extracolumella, red = quadrate, green = lower jaw. Scale bars = 1 mm.

Bellairs, 1953; Manley & Kraus, 2010; Daza & Bauer, 2015), but these modifications have not been reported in Pletholax gracilis or Ophidiocephalus taeniatus. The tympanic pygopodids have dorsoventrally inflected quadrates that are characterized by a large posterior concavity and lateral crest, whereas the non-tympanic pygopodids possess columnar quadrates that are mediolaterally compressed and lack a posterior concavity and lateral expansion (Figs 3E-I, S2). Daza & Bauer (2015) suggested that Aprasia repens may possess poor sound-pressure detection but is sensitive to substrate vibrations if the modified quadrate is able to act as a structural link in transmitting low-frequency vibrations from the jaw to the inner ear. We hypothesize that Aprasia, Pletholax and Ophidiocephalus all utilize their quadrate for this function due to their anatomical similarities in quadrate structure, reductions in ear morphology, and tight articulation between the quadrate and paroccipital process. These miniaturized non-tympanic pygopodids are burrowing species (Rieppel, 1985; Shea & Peterson, 1993; Cogger, 2014); therefore, a shift from sound-pressure detection to substrate vibration sensitivity would be advantageous (Christensen et al., 2012).

Non-tympanic reception of substrate vibrations has been confirmed in other squamates, including in snakes and amphisbaenians (Gans & Wever, 1972; Wever, 1979; Christensen et al., 2012). However, the structural link between the lower jaw and inner ear appears to be highly variable in these non-tympanic taxa (Fig. 7). The stapes in snakes are generally anchored to the quadrate via one or more intervening cartilages (McDowell, 1967; Young, 2015; Fig. 7C), whereas in most amphisbaenians, an elongated stapes and extracolumella extend anteriorly, bypassing the quadrate, and contacting the lateral surface of the lower jaw directly (Wever, 1979; Fig. 7B). The non-tympanic pygopodids appear to be unique in reducing the size and function of the stapes and extracolumella as part of the structural link and may utilize an alternative, quadrate-only mechanism because of the close association between the quadrate, paroccipital process and inner ear (Figs 7A, S2). These different anatomical mechanisms may be explained by alternative foraging strategies. Aprasia, Pletholax and Ophidiocephalus primarily forage on very small invertebrates, such as ants and termites, and their larvae (Patchell & Shine, 1986b; Shea & Peterson, 1993; Webb & Shine 1994), which probably require little cranial kinesis (including little streptostyly) and a relatively small gape, permitting the quadrate to be shifted anteriorly and abut the otic capsule and inner ear. Macrostomatan snakes generally forage on very large prey, utilizing extreme cranial kinesis capabilities and a large gape, thereby constraining the quadrate in a posterior position, suspended from

an extended supratemporal (Gans, 1961; Fig. 7C). The quadrate in snakes is distant from the inner ear and requires the structural link of the stapes. However, a number of snake species that occupy underground macrohabitats (e.g. leptotyphlopids, anomalepidids, uropeltids, some colubroids) possess an anteriorly shifted quadrate, a reduced stapedial shaft (but large footplate) and an insectivorous diet (Rieppel, 1979; Rieppel *et al.*, 2009; Olori & Bell, 2012; Daza & Bauer, 2015, Scanferla, 2016), suggesting that many fossorial squamate lineages have independently evolved these traits.

CONCLUSIONS

Our study has highlighted the importance of considering multiple factors that may influence the evolution of morphological phenotypes. Although we emphasized the gekkotan taxa that demonstrate particularly strong influences of phylogenetic history, allometry or potential functional demands, most taxa exhibit intermediate forms that have likely been influenced by the interaction of these three factors, as well as others that were not explored in this study. It is also important to note that the quadrate is a single functional unit within an integrated skull and studying the mechanisms of diversification in the remaining cranial elements will provide a more comprehensive understanding of gekkotan skull evolution as a whole. The evolution of the quadrate bone remains understudied in all squamates, and its role in jaw mechanics, cranial kinesis and support of the auditory system requires further investigation in a comparative and evolutionary framework.

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REFERENCES

- **Adams DC. 2014a.** A generalized K statistic for estimating phylogenetic signal from shape and other high-dimensional multivariate data. *Systematic Biology* **63:** 685–697.
- **Adams DC. 2014b.** A method for assessing phylogenetic least squares models for shape and other high-dimensional data. *Evolution* **68:** 2675–2688.
- Adams DC, Otarola-Castillo E. 2013. geomorph: an R package for the collection and analysis of geometric morphometric shape data. Methods in Ecology and Evolution 4: 393–399.
- Angelici FM, Luiselli L, Rugiero L. 1997. Food habits of the green lizard, *Lacerta bilineata*, in central Italy and a reliability test of faecal pellet analysis. *Italian Journal of Zoology* 64: 443–454.
- Bauer AM. 1986. Systematics, biogeography and evolutionary morphology of the Carphodactylini (Reptilia: Gekkonidae). Unpublished Ph.D. Dissertation, University of California, Berkeley.
- Bauer AM. 1990a. Gekkonid lizards as prey of invertebrates and predators of vertebrates. *Herpetological Review* 21: 83–87.
- Bauer AM. 1990b. Phylogenetic systematics and biogeography of the Carphodactylini (Reptilia: Gekkonidae). Bonner Zoologische Monographien 30: 1–218.
- Bauer AM, Sadlier RA. 1994. Diet of the new Caledonian gecko Rhacodactylus auriculatus (Squamata, Gekkonidae). Russian Journal of Herpetology 1: 108–113.
- **Blomberg SP, Garland T, Ives AR. 2003.** Testing for phylogenetic signal in comparative data: behavioral traits are more labile. *Evolution* **57:** 717–745.
- **Bookstein FL. 1991.** Morphometric tools for landmark data: geometry and biology. Cambridge: Cambridge University Press.
- Christensen CB, Christensen-Dalsgaard J, Brandt C, Madsen PT. 2012. Hearing with an atympanic ear: good vibration and poor sound-pressure detection in the royal python, *Python regius*. The Journal of Experimental Biology 215: 331–342.
- Cogger HG. 2014. Reptiles and amphibians of Australia, 7th edn. Clayton: CSIRO Publishing.
- Collyer ML, Sekora DJ, Adams DC. 2015. A method for analysis of phenotypic change for phenotypes described by high-dimensional data. *Heredity* 115: 357–365.
- Daza JD, Abdala V, Thomas R, Bauer AM. 2008. Skull anatomy of the miniaturized gecko Sphaerodactylus

- roosevelti (Squamata: Gekkota). Journal of Morphology **269:** 1340–1364.
- Daza JD, Bauer AM. 2012. Temporal bones of the Gekkota support molecular relationships within the Pygopodoidea. Journal of Herpetology 46: 381–386.
- Daza JD, Bauer AM. 2015. Cranial anatomy of the pygopodid lizard Aprasia repens, a gekkotan masquerading as a Scolecophidian. In: Emonds ORP, Powell GL, Jamniczky HA, Bauer AM, Theordor J, eds. All animals are interesting: a Festschrift in honour of Anthony P. Russell. Oldenburg: BIS-Verlag der Carl von Ossietzky Universität, 303–350.
- Daza JD, Diogo R, Johnston R, Abdala V. 2011. Jaw adductor muscles across lepidosaurs: a reappraisal. The Anatomical Record 294: 1765-1782.
- Daza JD, Herrera A, Thomas R, Claudio HJ. 2009. Are you what you eat? A geometric morphometric analysis of gekkotan skull shape. *Biological Journal of the Linnean Society* 97: 677–707
- Doughty P, Shine R. 1995. Life in two dimensions: natural history of the southern leaf-tailed gecko, *Phyllurus platurus*. *Herpetologica* 51: 193–201.
- **Drake AG, Klingenberg CP. 2008.** The pace of morphological change: historical transformation of skull shape in St. Bernard dogs. *Proceedings of the Royal Society of London B* **275:** 71–76.
- Drake AG, Klingenberg CP. 2010. Large-scale diversification of skull shape in domestic dogs: disparity and modularity. The American Naturalist 175: 289–301.
- Earle AM. 1961. The middle ear of *Holbrookia* and *Callisaurus*. Copeia 1961: 405–410.
- **Evans SE. 2008.** The skull of lizards and tuatara. In: Gans C, Gaunt AS, Adler K, eds. *Biology of the Reptilia, morphology H: the skull of Lepidosauria. Volume 20.* New York: Society for the Study of Amphibians and Reptiles, 1–344.
- Felice RN, Goswami A. 2017. Developmental origins of mosaic evolution in the avian cranium. *Proceedings of the National Academy of Sciences of the United States of America* 115: 555–560.
- Frazzetta TH. 1962. A functional consideration of cranial kinesis in lizards. *Journal of Morphology* 111: 287–319.
- Gamble T, Greenbaum E, Jackman TR, Bauer AM. 2015. Into the light: diurnality has evolved multiple times in geckos. *Biological Journal of the Linnean Society* 115: 896–910.
- Gamble T, Greenbaum E, Jackman TR, Russell AP, Bauer AM. 2012. Repeated origin and loss of adhesive toepads in geckos. PLoS ONE 7: e39429.
- **Gans C. 1961.** The feeding mechanism of snakes and its possible evolution. *American Zoologist* 1: 217–227.
- Gans C. 1986. Evolution of limbless squamates: functional aspects. In: Rocek Z, ed. Studies in herpetology. Prague: Charles University Press, 71–74.
- Gans C, Wever EG. 1972. The ear and hearing in Amphisbaenia (Reptilia). *Journal of Experimental Zoology* 179: 17–34.
- Goswami A, Polly PD. 2010. The influence of modularity on cranial morphological disparity in Carnivora and Primates (Mammalia). PLoS ONE 5: e9517.
- **Gould SJ, Lewontin RC. 1979.** The spandrels of San Marco and the Panglossian paradigm: a critique of the adaptationist

- programme. Proceedings of the Royal Scoiety of London B **205**: 581–598.
- Grant PR, Grant BR. 2002. Unpredictable evolution in a 30-year study of Darwin's Finches. Science 296: 707-711.
- Herrel A, Aerts P, de Vree F. 2000. Cranial kinesis in geckoes: functional implications. The Journal of Experimental Biology 203: 1415–1423.
- Herrel A, de Grauw E, Lemos-Espinal JA. 2001. Head shape and bite performance in xenosaurid lizards. *Journal of Experimental Zoology* 290: 101–107.
- Herrel A, de Vree F, Delheusy V, Gans C. 1999. Cranial kinesis in gekkonid lizards. The Journal of Experimental Biology 202: 3687–3698.
- Herrel A, Joachim R, Vanhooydonck B, Irschick DJ. 2006. Ecological consequences of ontogenetic changes in head shape and bite performance in the Jamaican lizard Anolis lineatopus. Biological Journal of the Linnean Society 89: 443–454.
- Herrel A, O'Reilly JC. 2006. Ontogenetic scaling of bite force in lizards and turtles. *Physiological and Biochemical Zoology* 79: 31–42.
- Herrel A, Schaerlaeken V, Meyers JJ, Metzger KA, Ross CF. 2007. The evolution of cranial design and performance in squamates: consequences of skull-bone reduction on feeding behavior. *Integrative and Comparative Biology* 47: 107–117.
- Herrel A, Vanhooydonck B, Van Damme R. 2004. Omnivory in lacertid lizards: adaptive evolution or constraint? *Journal* of Evolutionary Biology 17: 974–984.
- How RA, Dell J, Wellington BD. 1990. Reproductive and dietary biology of Nephrurus and Underwoodisaurus (Gekkonidae) in Western Australia. Records of the Western Australian Museum 14: 449–459.
- **Lappin AK, Hamilton PS, Sullivan BK. 2006.** Bite-force performance and head shape in a sexually dimorphic crevice-dwelling lizard, the common chuckwalla [Sauromalus ater (= obesus)]. Biological Journal of the Linnean Society 88: 215–222.
- **Losos JB. 1990.** Ecomorphology, performance capability, and scaling of West Indian *Anolis* lizards: an evolutionary analysis. *Ecological Monographs*: **60**: 369–388.
- **Loveridge A. 1947.** Revision of the African lizards of the family Gekkonidae. *Bulletin of the Museum of Comparative Zoology* **98:** 1–469.
- Manley GA, Kraus JEM. 2010. Exceptional high-frequency hearing and matched vocalizations in Australian pygopod geckos. *Journal of Experimental Biology* 213: 1876–1885.
- McBrayer LD. 2004. The relationship between skull morphology, biting, performance, and foraging mode in Kalahari lacertid lizards. Zoological Journal of the Linnean Society 140: 403–416.
- McDowell SB. 1967. The extracolumella and tympanic cavity of the "earless" monitor lizard *Lanthanotus borneensis*. *Copeia* 1967: 154–159.
- Meiri S. 2008. Evolution and ecology of lizard body sizes. Global Ecology and Biogeography 17: 724–734.
- Metzger K. 2002. Cranial kinesis in lepidosaurs: skulls in motion. In: Aerts P, Herrel A, Van Damme R, eds. *Topics in functional and ecological vertebrate morphology*. Maastricht: Shaker Publishing, 15–46.

- Metzger K, Herrel A. 2005. Correlations between lizard cranial shape and diet: a quantitative, phylogenetically informed analysis. *Biological Journal of the Linnean Society* 86: 433–466.
- Meyers JJ, Herrel A, Birch J. 2002. Scaling of morphology, bite force, and feeding kinematics in an iguanian and a scleroglossan lizard. In: Aerts P, D'Août K, Herrel A, Van Damme R, eds. *Topics in functional and ecological vertebrate morphology*. Maastricht: Shaker Publishing, 47–62.
- Oelrich TM. 1956. The anatomy of the head of Ctenosaura pectinata (Iguanidae). Miscellaneous Publications of Zoology, University of Michigan 94: 1–122.
- Olori J, Bell CJ. 2012. Comparative skull morphology of uropeltid snakes (Alethinophidia: Uropeltidae) with special reference to disarticulated elements and variation. PLoS ONE 7: e32450.
- **Openshaw GH, Keogh JS. 2014.** Head shape evolution in monitor lizards (*Varanus*): interactions between extreme body size disparity, phylogeny and ecology. *Journal of Evolutionary Biology* **27:** 363–373.
- Paluh DJ, Olgun K, Bauer AM. 2018. Ontogeny, but not sexual dimorphism, drives the intraspecific variation of quadrate morphology in *Hemidactylus turcicus* (Squamata: Gekkonidae). *Herpetologica* 74: 22–28.
- Patchell FC, Shine R. 1986a. Feeding mechanisms in pygopodid lizards: how can *Lialis* swallow such large prey? *Journal* of *Herpetology* 20: 59–64.
- Patchell FC, Shine R. 1986b. Food habits and reproductive biology of the Australian legless lizards (Pygopodidae). *Copeia* 1986: 30–39.
- **Pianka ER, Huey RB. 1978.** Comparative ecology, resource utilization, and niche segregation among gekkonid lizards in the southern Kalahari. *Copeia* **1978:** 691–701.
- Revell LJ. 2012. Phytools: an R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution* 3: 217–223.
- **Rieppel O. 1978.** Streptostyly and muscle function in lizards. *Experientia* **34:** 776–777.
- Rieppel O. 1979. The braincase of Typhlops and Leptotyphlops (Reptilia: Serpentes). Zoological Journal of the Linnean Society 65: 161–176.
- Rieppel O. 1984. The structure of the skull and jaw adductor musculature in the Gekkota, with comments on the phylogenetic relationships of the Xantusiidae (Reptilia: Lacertilia). Zoological Journal of the Linnean Society 82: 291–318.
- Rieppel O. 1985. The recessus scalae tympani and its bearing on the classification of reptiles. *Journal of Herpetology* 19: 373–384.
- Rieppel O. 1993. Patterns of diversity in the reptilian skull. In: Hanken J, Hall BK, eds. *The skull, volume 2: patterns of structural and systematic diversity*. Chicago: University of Chicago Press, 344–390.
- Rieppel O, Kley NJ, Maisano JA. 2009. Morphology of the skull of the white-nosed blindsnake, *Liotyphlops albirostris* (Scolecophidia: Anomalepididae). *Journal of Morphology* 270: 536–557.
- **Rohlf FJ, Slice DE. 1990.** Extensions of the Procrustes method for the optimal superimposition of landmarks. *Systematic Zoology* **39:** 40–59.

- Rohtla EA. 2016. The call of the gecko: an acoustic and morphological examination of gecko vocalizations. Unpublished M.Sc. Thesis, Villanova University.
- Saenz D. 1996. Dietary overview of Hemidactylus turcicus with possible implications of food partitioning. Journal of Herpetology 30: 461–466.
- **Sanger TJ, Mahler DL, Abzhanov A, Losos JB. 2011.** Roles of modularity and constraint in the evolution of cranial diversity among *Anolis* lizards. *Evolution* **66:** 1525–1542.
- Scanferla A. 2016. Postnatal ontogeny and the evolution of macrostomy in snakes. Royal Society Open Science 3: 160612.
- Seilacher A. 1970. Arbeitskonzept zur Konstruktionsmorphologie. Lethaia 3: 393-396.
- **Shea GM, Peterson M. 1993.** Notes on the biology of the genus *Pletholax* Cope (Squamata: Pygopodidae). *Records of the Western Australian Museum* **16:** 419–425.
- **Shine R. 1986.** Evolutionary advantages of limblessness: evidence from the pygopodid lizards. *Copeia* **1986:** 525–529.
- Shute CCD, Bellairs Ad'A. 1953. The cochlear apparatus of Geckonidae and Pygopodidae and its bearing on the affinities of these groups of lizards. Proceedings of the Zoological Society London 123: 695–708.
- Snyder J, Synder L, Bauer AM. 2010. Ecological observations on the Gargoyle Gecko, *Rhacodactylus auriculatus* (Bavay, 1869), in southern New Caledonia. *Salamandra* 46: 37–47.
- Stephenson NG. 1960. The comparative osteology of Australian geckos and its bearing on their morphological status. Zoological Journal of the Linnean Society London 44: 278–299.
- **Underwood G. 1957.** On lizards of the family Pygopodidae. A contribution to the morphology and phylogeny of the Squamata. *Journal of Morphology* **100:** 207–268.

- Versluys J. 1912. Das Streptostylie problem und die Bewegungen im Schädel bei Sauropsiden. Zoologische Jahrbücher (Suppl.) 15: 545–716.
- Verwaijen D, Van Damme R, Herrel A. 2002. Relationships between head size, bite force, prey handling efficiency and diet in two sympatric lacertid lizards. Functional Ecology 16: 842–850.
- Webb JK, Shine R. 1994. Feeding habits and reproductive biology of Australian pygopodid lizards of the genus *Aprasia*. Copeia 1994: 390–398.
- **Webster DB. 1966.** Ear structure and function in modern mammals. *American Zoologist* **6:** 451–466.
- Werner YL, Igíc PG. 2002. The middle ear of gekkonoid lizards: interspecific variation of structure in relation to body size and to auditory sensitivity. *Hearing Research* 167: 33–45.
- Werner YL, Igíc PG, Seifan M, Saunders JC. 2002. Effects of age and size in the ears of gekkonomorph lizards: middle-ear sensitivity. *The Journal of Experimental Biology* **205**: 3215–3223.
- Werner YL, Safford SD, Seifan M, Saunders JC. 2005. Effects of age and size in the ears of gekkonomorph lizards: middle-ear morphology with evolutionary implications. *The Anatomical Record Part A*: 283: 212–223.
- Wever EG. 1973. Closure muscles of the external auditory meatus in Gekkonidae. *Journal of Herpetology* 7: 323–329.
- Wever EG. 1979. The reptile ear. Princeton: Princeton University Press.
- Whitfield SM, Donnelly MA. 2006. Ontogenetic and seasonal variation in the diets of a Costa Rican leaf-litter herpetofauna. *Journal of Tropical Ecology* 22: 409–417.
- Young BA. 2015. The evolution of the snake ear and its restricted frequency response range. In: Emonds ORP, Powell GL, Jamniczky HA, Bauer AM, Theordor J, eds. *All animals are interesting: a Festschrift in honour of Anthony P. Russell*. Oldenburg: BIS-Verlag der Carl von Ossietzky Universität, 423–437.

SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site.

- **Figure S1.** Phylomorphospace plots of gekkotan quadrate shape variation. Black points are specimens projected into tangent space and white points are estimated ancestral states.
- **Figure S2.** Posterior view of quadrate (left) and lateral view of skull (right). A, *Paradelma orientalis* (CAS 77652); B, *Aprasia repens* (CAS 104382); C, *Pletholax gracilis* (MCZ R-187676); D, *Ophidiocephalus taeniatus* (AM R-45179). E, Phylomorphspace plot illustrating the divergence in quadrate shape between tympanic pygopodids (orange) and nontympanic pygopodids (purple). Quadrates in phylomorphospace plot are in lateral view.
- **Figure S3.** Labelled PCA plot of gekkotan quadrates in tangent space (see Table S1 for plot numbers). See colour designations in Figure 4.
- **Figure S4.** Labelled multivariate regression plot between quadrate shape and quadrate centroid size (see Table S1 for plot numbers). See colour designations in Figure 4.
- **Table S1.** Gekkotan specimens examined in this study and data associated with them. Institutional abbreviations for the specimens studied are as follows: AMNH, American Museum of Natural History; AMS, Australian Museum; CAS, California Academy of Sciences; CM, Carnegie Museum of Natural History; FMNH, Field Museum of Natural History; MCZ, Museum of Comparative Zoology, Harvard University; QM, Queensland Museum; SAMA, South Australian Museum; UMMZ, University of Michigan Museum of Zoology; USNM, National Museum of Natural History, USA; YPM, Yale Peabody Museum of Natural History.
- **Table S2.** Kappa statistic and *P* value associated with phylogenetic signal assessment. Groups with phylogenetic signal approaching significance are marked with an asterisk (*).